# Synthesis and Serotonergic Activity of N,N-Dimethyl-2-[5-(1,2,4-triazol-1-ylmethyl)-1H-indol-3-yl]ethylamine and Analogues: Potent Agonists for 5-HT<sub>1D</sub> Receptors<sup>1</sup>

Leslie J. Street,\*,<sup>†</sup> Raymond Baker,<sup>†</sup> William B. Davey,<sup>†</sup> Alexander R. Guiblin,<sup>†</sup> Richard A. Jelley,<sup>†</sup> Austin J. Reeve,<sup>†</sup> Helen Routledge,<sup>†</sup> Francine Sternfeld,<sup>†</sup> Alan P. Watt,<sup>†</sup> Margaret S. Beer,<sup>‡</sup> Derek N. Middlemiss,<sup>‡</sup> Alison J. Noble,<sup>‡</sup> Josephine A. Stanton,<sup>‡</sup> Kate Scholey,<sup>§</sup> Richard J. Hargreaves,<sup>§</sup> Bindi Sohal,<sup>∇</sup> Michael I. Graham,<sup>∇</sup> and Victor G. Matassa<sup>†</sup>

Chemistry, Biochemistry, and Pharmacology Departments, Merck Sharp & Dohme Research Laboratories, Neuroscience Research Centre, Terlings Park, Eastwick Road, Harlow, Essex CM20 2QR, U.K., and Predevelopment Pharmacokinetic Department, Merck Sharp & Dohme Research Laboratories, Development Laboratories, Hertford Road, Hoddesdon, Hertfordshire EN11 9BU, U.K.

Received January 26, 1995<sup>∞</sup>

The synthesis and the 5-HT receptor activity of a novel series of  $N_{N}$ -dimethyltryptamines substituted in the 5-position with an imidazole, triazole, or tetrazole ring are described. The objective of this work was to identify potent and selective 5-HT<sub>1D</sub> receptor agonists with high oral bioavailability and low central nervous system penetration. Compounds have been prepared in which the azole ring is attached through either nitrogen or carbon to the indole. Conjugated and methylene-bridged derivatives have been studied (n = 0 or 1). Substitution of the azole ring has been explored either  $\alpha$  or  $\beta$  to the point of attachment to indole. In a series of N-linked azoles (X = N), simple unsubstituted compounds have high affinity and selectivity for 5-HT<sub>1D</sub> receptors. It is proposed that for good affinity and selectivity a hydrogen bond acceptor interaction with the 5-HT<sub>1D</sub> receptor, through a  $\beta$ -nitrogen in the azole ring, is required. In a series of C-linked triazoles and tetrazoles (X = C), optimal affinity and selectivity for the 5-HT<sub>1D</sub> receptor was observed when the azole ring is substituted at the 1-position with a methyl or ethyl group. This study has led to the discovery of the 1,2,4-triazole 10a (MK-462) as a potent and selective 5-HT<sub>1D</sub> receptor agonist which has high oral bioavailability and rapid oral absorption. The *in vitro* activity and the preliminary pharmacokinetics of compounds in this series are presented.

# Introduction

During the last 5 years, molecular biology has revealed the immense diversity of serotonin (5-HT, 1) (Chart 1) receptors. At this point in time, seven 5-HT receptor families have been identified of which 5-HT<sub>1,2,4-7</sub> are G-protein-coupled receptors.<sup>2-8</sup> For many of these 5-HT families, receptor subtypes have been identified and classified on the basis of a combination of amino acid sequence homology in the seven transmembrane domains, the signal transduction mechanism, and classical pharmacology.<sup>9</sup> The most recently cloned subtypes hold promise for the discovery of new selective drug candidates in the next 5-10 years. The 5-HT<sub>1</sub> family appears to have the highest multiplicity, and to date five human 5-HT<sub>1</sub>-like receptors have been cloned, 5-HT<sub>1A</sub>, 5-HT<sub>1B/1D $\beta$ </sub>, 5-HT<sub>1D $\alpha$ </sub>, 5-HT<sub>1E</sub>, and 5-HT<sub>1F</sub>.<sup>2,3</sup> Much work recently has focused on 5-HT<sub>1D</sub> receptors,<sup>10</sup> originally characterized in bovine brain membranes<sup>11</sup> and shown to have high affinity for the 5-HT<sub>1</sub> selective agonist 5-carbamoyltryptamine (5-CT, 2).12 The introduction of the 5-HT<sub>1D</sub> receptor agonist sumatriptan (3), for the acute treatment of migraine,<sup>13</sup> has sparked an intense research effort to discover more potent and selective 5-HT<sub>1D</sub> receptor agonists with improved pharmacokinetic profiles.<sup>14-18</sup> Sumatriptan selectively constricts intracranial vascular smooth muscle and inhibits neuropeptide release from perivascular trigeminal sensory neurones, and both mechanisms have been proposed to be important in eliciting its antimigraine action.<sup>19,20</sup>

We have recently reported that the carboxamide and sulfonamide groups of 2 and 3, respectively, can be replaced with substituted 1,2,4-oxadiazole and 1,2,4thiadiazole rings to give potent and selective 5-HT<sub>1D</sub> receptor agonists, and we concluded that the H-bond acceptor ability of these rings was important for 5-HT<sub>1D</sub> receptor affinity and selectivity.<sup>21,22</sup> Structure-activity studies in these series led to the discovery of the benzylsulfonamide 4a (L-694,247) as a highly potent 5-HT<sub>1D</sub> receptor agonist with good selectivity for 5-HT<sub>1D</sub> receptors.<sup>23</sup> Pharmacokinetic studies on compounds such as 4a, however, suggested low oral bioavailability. In the oxadiazole series, it was generally found that primary tryptamines had low oral bioavailability, probably as a result of metabolism by monoamine oxidase. The N,N-dimethyltryptamine 4b (L-695,894) however, showed 40% oral bioavailability in rats but had significant affinity for 5-HT<sub>2A</sub> and 5-HT<sub>2C</sub> receptors.<sup>21</sup> In order to improve the 5-HT<sub>1D</sub> receptor selectivity and oral bioavailability, we have extended this work to explore alternative 5-membered heteroaromatic rings which are also capable of functioning as H-bond acceptors. Compounds were sought which had log D's < -0.5 to minimize central nervous system (CNS) penetration.<sup>13</sup> We describe herein the synthesis, serotonergic activity, and preliminary pharmacokinetics of a series of N,N-

<sup>&</sup>lt;sup>†</sup> Chemistry Department. <sup>‡</sup> Biochemistry Department.

<sup>&</sup>lt;sup>§</sup> Pharmacology Department.

<sup>&</sup>lt;sup>v</sup> Predevelopment Pharmacokinetic Department.

<sup>&</sup>lt;sup>®</sup> Abstract published in Advance ACS Abstracts, May 1, 1995.

Chart 1



<sup>a</sup> Reagents: (a) NaH, DMF, 4-nitrobenzyl bromide; (b) H<sub>2</sub>, 10% Pd-C, EtOH, 2 N HCl; (c) NaNO<sub>2</sub>, H<sub>2</sub>O, concentrated HCl, -10 °C; (d) SnCl<sub>2</sub>·2H<sub>2</sub>O, concentrated HCl; (e) NaOH (aq)/EtOAc; (f) 4-chlorobutanal dimethyl acetal, EtOH/H<sub>2</sub>O (5:1), 5 N HCl (1.2 equiv), reflux, 4 h; (g) HCHO, NaCNBH<sub>3</sub>, MeCO<sub>2</sub>H, MeOH, 0 °C; (h) 4-(N,N-dimethylamino)butanal dimethyl acetal, 4% H<sub>2</sub>SO<sub>4</sub>, reflux, 2 h.

10a-c

dimethyltryptamines (5), substituted in the 5-position of indole with imidazole, triazole, or tetrazole and linked through either N or C in the ring. The choice of these heterocycles also allowed a study of the effect of substitution in the ring, either  $\alpha$  or  $\beta$  to the point of attachment to indole, on 5-HT<sub>1D</sub> receptor affinity and selectivity. This work has led to the discovery of the 1,2,4-triazole 10a (L-705,126, MK-462) as a potent and selective 5-HT<sub>1D</sub> receptor agonist with high oral bioavailability and good in vivo activity predictive of antimigraine action.<sup>24</sup>

### Synthetic Chemistry

The imidazole-, triazole-, and tetrazole-substituted N,N-dimethyltryptamines 10a-c and 14a-d were prepared starting from 4-nitrobenzyl bromide (Schemes 1

and 2). Reaction of the sodium derivative of 1,2,4triazole with 4-nitrobenzyl bromide gave 4-nitrobenzyl triazole 7a as a crystalline solid and, as expected, as a single isomer<sup>25a</sup> (Scheme 1). Similar treatment of 4-nitrobenzyl bromide with imidazole and 2-methylimidazole gave the 4-nitrobenzyl imidazoles 7b,c, respectively. Alkylation of 1-H-1,2,3-triazole with 4-nitrobenzyl bromide gave a 6:1 mixture of 11a:b which were separated by silica gel chromatography (Scheme 2). Reaction of 1-H-tetrazole with 4-nitrobenzyl bromide under the same conditions gave the alkylation products 11c (75%) and 11d (17%). The regiochemical assignments for 11a-d were made on the basis of NOE enhancement experiments. Saturation of the H-5 imidazole and tetrazole protons in 11a,c, respectively, gave an NOE enhancement of CH<sub>2</sub> thus defining N-1 as the

9a-c

#### Scheme $2^a$



<sup>a</sup> Reagents: (a) NEt<sub>3</sub>, MeCN, 4-nitrobenzyl bromide; (b) H<sub>2</sub>, 10% Pd-C, EtOH, 2 N HCl; (c) NaNO<sub>2</sub>, H<sub>2</sub>O, concentrated HCl, -10 °C; (d) SnCl<sub>2</sub>·2H<sub>2</sub>O, concentrated HCl; (e) NaOH (aq)/EtOAc; (f) 4-chlorobutanal dimethyl acetal, EtOH/H<sub>2</sub>O (5:1), 5 N HCl (1.2 equiv), reflux, 4 h; (g) HCHO, NaCNBH<sub>3</sub>, MeCO<sub>2</sub>H, MeOH, 0 °C.

#### Scheme 3<sup>a</sup>



<sup>a</sup> Reagents: (a) NaH, DMF, 4-fluoronitrobenzene; (b) H<sub>2</sub>, 10% Pd-C, EtOH, 2 N HCl; (c) NaNO<sub>2</sub>, H<sub>2</sub>O, concentrated HCl, -10 °C; (d) SnCl<sub>2</sub>·2H<sub>2</sub>O, concentrated HCl; (e) NaOH (aq/EtOAc; (f) 4-chlorobutanal dimethyl acetal, EtOH/H<sub>2</sub>O (5:1), 5 N HCl (1.2 equiv), reflux, 4 h; (g) HCHO, NaCNBH<sub>3</sub>, MeCO<sub>2</sub>H, MeOH, 0 °C.

position of alkylation of the heterocycles. Hydrogenation of 7a-c and 11a-d over Pd-C gave the anilines 8a-c and 12a-d which were isolated as their hydrochloride salts. Treatment of 8a-c and 12a-d with NaNO<sub>2</sub> followed by reduction of the intermediate diazonium salts with SnCl<sub>2</sub>·2H<sub>2</sub>O gave the hydrazines 9a-c and 13a-d. Fischer reaction<sup>25b</sup> of 9a-c and 13a-d with 4-chlorobutanal dimethyl acetal,<sup>26</sup> in refluxing EtOH/H<sub>2</sub>O (5:1) and 5 N HCl (1.2 equiv), afforded the corresponding tryptamines which were treated with NaCNBH<sub>3</sub>/CH<sub>2</sub>O/MeCO<sub>2</sub>H to give the N,Ndimethyltryptamines 10a-c and 14a-d respectively, in moderate yields (Schemes 1 and 2).<sup>27</sup> An alternative procedure for the preparation of triazole 10a was to treat the hydrochloride salt of hydrazine 9a with 4-(N,Ndimethylamino)butanal dimethyl acetal,<sup>28</sup> in refluxing 4% sulfuric acid, to give N,N-dimethyltryptamine 10a directly.

The route to the conjugated triazole **18a** and imidazole **18b** is shown in Scheme 3. Reaction of the sodium salt of 1,2,4-triazole with 1-fluoro-4-nitrobenzene, in DMF, gave a high yield of the N-1 alkylation product **15a** together with a trace amount of the N-4 adduct. Similar reaction of 2-methylimidazole with 1-fluoro-4nitrobenzene gave **15b**. Hydrogenation of **15a,b** gave the anilines **16a,b** which were converted to the hydrazines 17a,b using NaNO<sub>2</sub>/SnCl<sub>2</sub>·2H<sub>2</sub>O. Fischer reaction of 17a,b with 4-chlorobutanal dimethyl acetal followed by N,N-dimethylation of the resultant tryptamines afforded 18a,b.

The N-methyl-1,2,4-triazoles **21a,b** and **26** were prepared as illustrated in Schemes 4 and 5. The imino ether 20 was prepared by treatment of N,N-dimethyl-2-[5-(cyanomethyl)-1H-indol-3-yl]ethylamine, 19,<sup>21</sup> with EtOH/HCl (gas). Treatment of 20 with methylhydrazine followed by refluxing in formic acid gave the N-methyl-1,2,4-triazoles **21a,b** in a 1:2 ratio, respectively, in low yield. Regiochemical assignments were again based on NOE enhancement experiments. Saturation of the methyl of **21a**, **b** gave NOE enhancements of the triazole proton and methylene bridge, respectively, thus defining the regiochemistry. The 4-methyl-1,2,4-triazole **26** was prepared starting from the methyl imino ether 22 (Scheme 5). Addition of formylhydrazine to 22 gave the formylhydrazone 23 which was reacted with methylamine to give the desired 4-methyl-1,2,4triazole 24, in 87% yield, after silica gel chromatography. Hydrogenation followed by diazotization and reduction gave the hydrazine 25. Fischer reaction of 25 with 4-(N,N-dimethylamino) but anal dimethyl acetal, in refluxing 4% H<sub>2</sub>SO<sub>4</sub>, gave **26** in moderate yield.

#### Scheme $4^a$



<sup>a</sup> Reagents: (a) EtOH/HCl; (b) MeNHNH<sub>2</sub>, EtOH, NEt<sub>3</sub>; (c) HCO<sub>2</sub>H, reflux, 2 h.

Scheme  $5^a$ 



<sup>a</sup> Reagents: (a) NH<sub>2</sub>NHCHO, MeOH; (b) MeNH<sub>2</sub> (g), MeOH; (c) H<sub>2</sub>, 10% Pd-C, EtOH, 2 N HCl; (d) NaNO<sub>2</sub>, H<sub>2</sub>O, concentrated HCl, -10 °C; (e) SnCl<sub>2</sub>·2H<sub>2</sub>O, concentrated HCl; (f) 4-(N,N-dimethylamino)butanal dimethyl acetal, 4% H<sub>2</sub>SO<sub>4</sub>, reflux, 2 h.

A series of tetrazoles substituted on either N-1 or N-2 was prepared (Scheme 6). The tetrazole ring of  $N_{,N}$ dimethyltryptamine 28 and tryptamine 29 was constructed by reaction of the corresponding nitriles 19 and 27, respectively, with sodium azide, in N-methyl-2pyrrolidinone, using triethylamine hydrochloride as catalyst. Tetrazoles 28 and 29 were obtained in 76% and 69% yield, respectively. Tetrazole formation was not observed when the free bases of 19 and 27 were used. In order to alkylate the tetrazole ring of 29, protection of the tryptamine as the Boc derivative 30 was necessary. Reaction of 30 with methyl iodide gave a mixture of the N-1 and N-2 alkylation products 31a,b in a 1.9:1 ratio which were separated by silica gel chromatography. Similar reaction of 30 with ethyl iodide gave the ethyl tetrazoles **33a,b** in 43% and 37% yields, respectively. In each case, the N-1 alkylation products were more polar than the N-2 adducts on silica gel and the structural assignments were made by NOE enhancement experiments. Thus, saturation of the methyl of **31a** and the  $CH_2CH_3$  of **33a** gave NOE enhancement of the methylene bridge in each compound, thus defining the position of alkylation as N-1. Removal of the Boc group of 31a,b and 33a,b was achieved using trifluoroacetic acid to give the tryptamines 32a,b and 34a,b, respectively. Reaction of 32a,b and 34a,b with NaCNBH<sub>3</sub>/HCHO/CH<sub>3</sub>CO<sub>2</sub>H in MeOH gave the N,N-dimethyltryptamines **35a,b**, and **36a,b** respectively (Scheme 6).

## **Results and Discussion**

**Structure**-Affinity Relationships. The 5- $HT_{1D}$  receptor affinities of compounds were measured by displacement of [<sup>3</sup>H]-5-HT from bovine caudate membranes, in the presence of cyanopindolol and mesulergine to block interactions with 5- $HT_{1A}$ , 5- $HT_{1B}$ , and 5- $HT_{2C}$  sites.<sup>11,21,29</sup> The data is presented in Tables 1–3.

The 5-HT<sub>1D</sub> receptor affinities of a series of imidazoles, triazoles, and tetrazoles, attached to indole through a ring N, are shown in Table 1. The 1-substituted 1.2.4triazole 10a has good affinity for the 5-HT<sub>1D</sub> receptor and is comparable to the 1-substituted 1,2,3-triazole **14a.** Both compounds have low log D's, predictive of low CNS penetration (cf. sumatriptan log D = -1.17) which is important for potential antimigraine agents of this class.<sup>13a,b</sup> The 2-substituted 1,2,3-triazole 14b has reduced affinity compared to 10a and 14a, suggesting that a ring N,  $\beta$  to the position of substitution (X or Y = N), is important for binding to the 5-HT<sub>1D</sub> receptor in this series. The 4-substituted 1,2,4-triazole analogue of 10a could not be prepared because of stability problems. Imidazole 10b and tetrazoles 14c,d have comparable 5-HT<sub>1D</sub> receptor affinities to triazole 10a although  $\log D$  is optimal for 10a reflecting the higher

## Scheme 6<sup>a</sup>



<sup>a</sup> Reagents: (a) NaN<sub>3</sub>, N-methyl-2-pyrrolidinone, Et<sub>3</sub>N·HCl, 160 °C, 8–16 h; (b) (Boc)<sub>2</sub>O, NEt<sub>3</sub>, CH<sub>2</sub>Cl<sub>2</sub>; (c) R<sup>2</sup>I, NEt<sub>3</sub>, MeCN; (d) trifluoroacetic acid, CH<sub>2</sub>Cl<sub>2</sub>; (e) HCHO, NaCNBH<sub>3</sub>, MeCO<sub>2</sub>H, MeOH, 0 °C.



<sup>a</sup> SEM = standard error of the mean from  $n \ge 3$ . Where SEM is not quoted, the figures are the mean of two independent determinations typically with individual values within  $\pm (10-15)\%$  of the mean. <sup>b</sup> log P measured at pH 7.4.

basicity of this system. 2-Methyl substitution of the imidazole ring, to give **10c**, resulted in slightly lower affinity for 5-HT<sub>1D</sub> receptors compared to the unsubstituted analogue **10b**. The effect of conjugation of the 5-ring heterocycle with indole on 5-HT<sub>1D</sub> receptor affinity, in this series, was demonstrated by preparation of triazole **18a** and 2-methylimidazole **18b**. Both com-

pounds have 5-10-fold higher affinity for the 5-HT<sub>1D</sub> receptor than the methylene-linked analogues **10a**,c. This result parallels that we reported for a series of 3-substituted 1,2,4-oxadiazoles which led to the identification of the highly potent 5-HT<sub>1D</sub> receptor agonist L-694,247.<sup>21,23</sup> Unfortunately, a second consequence of conjugation is increased hydrophobicity, e.g., compare **10c**, log D = -0.74, and **18b**, log D = -0.13, resulting in greater penetration into the CNS.

The three regioisomers of N-methyl-1,2,4-triazole, **21a,b** and **26**, were prepared to determine the effect of changing the position of methyl substitution on 5-HT<sub>1D</sub> receptor binding and log D (Table 2). The 1,3-substituted triazole **21a** has 10-fold lower affinity for 5-HT<sub>1D</sub> receptors compared with sumatriptan. One order of magnitude improvement in activity was seen for the 4-methyltriazole **26**, whereas the 1,5-substituted triazole **21b** has intermediate 5-HT<sub>1D</sub> receptor affinity. Triazole **26** also has optimal hydrophilicity, again reflecting the higher basicity of this pattern of substitution in the 1,2,4-triazole ring.

Data for an analogous series of substituted tetrazoles are shown in Table 3. Here both the position of substitution in the ring and the size of the substituent were studied. The zwitterionic 1*H*-tetrazole **28** has poor affinity for the 5-HT<sub>1D</sub> recognition site suggesting that charged fragments at this position are not well tolerated by the receptor. Alkylation of the tetrazole ring gave up to 100-fold improvement in receptor affinity. In a series of methyl- and ethyl-substituted tetrazoles, the highest 5-HT<sub>1D</sub> receptor affinity was observed for the 1-substituted derivatives. Thus, 1-methyltetrazole **35a** has 5-fold higher affinity than the 2-methyl analogue **35b**. **Table 2.** Displacement of  $[^{3}H]$ -5-HT Binding to 5-HT<sub>1D</sub> Recognition Sites in Pig Caudate Membranes by C-Linked Triazoles



<sup>*a,b*</sup> See corresponding footnotes of Table 1.

**Table 3.** Displacement of  $[^{3}H]$ -5-HT Binding to 5-HT<sub>1D</sub> Recognition Sites in Pig Caudate Membranes by C-Linked Tetrazoles



<sup>*a,b*</sup> See corresponding footnotes of Table 1.

The ethyltetrazoles **36a**,**b** have comparable activity to the methyl derivatives **35a**,**b**, respectively. Structureactivity in the tetrazole series therefore parallels that seen in the methyl triazole series with substitution of the heteroaromatic ring at the  $\alpha$ -position leading to higher affinity compounds.

The 5-HT<sub>1D</sub> binding results for the N-linked imidazoles, triazoles, and tetrazoles 10a-c, 14a-d, and 18a.b (Table 1) demonstrate that unsubstituted 5-membered heteroaromatic rings are well tolerated at the 5-HT<sub>1D</sub> recognition site. It is predicted that the critical role of the azole ring at the receptor is to act as a H-bond acceptor. The lower affinity of the 2-substituted 1.2.3triazole 14b would suggest that this interaction is primarily through the  $\beta$ -nitrogen (X or Y in Table 1) of the azole ring in compounds 10a-c, 14a,c,d, and 18a,b. The importance of this interaction is also reflected in the results for the C-linked 1,2,4-triazoles and tetrazoles. The 5-10-fold higher affinity of 4-methyltriazole 26 and 1-methyltetrazole 35a, compared to the triazole 21a and 2-methyltetrazole 35b, can also be rationalized on the basis of the degree to which the  $\beta$ -nitrogen in the azole ring can participate in a H-bond interaction. This would be more favorable for compounds **26** and **35a**. However, in these systems a more favorable hydrophobic interaction of the receptor with the  $\alpha$ -methyl substituent cannot be ruled out.

**Receptor Selectivity**. The pharmacological specificity of the compounds for 5-HT<sub>1D</sub> receptors was determined by measuring affinities for 5-HT<sub>1A</sub>, 5-HT<sub>2A</sub>, 5-HT<sub>2C</sub>, and 5-HT<sub>3</sub> receptors. The data are presented in Table 4. The 1,2,4-triazole 10a has good selectivity for 5-HT<sub>1D</sub> receptors over 5-HT<sub>1A</sub> receptors (6-fold) and 5-HT<sub>2A</sub>, 5-HT<sub>2C</sub>, and 5-HT<sub>3</sub> receptors (all  $\geq$ 100-fold) and has an improved selectivity profile over the aminooxadiazole 4b. Imidazoles 10b,c have comparable selectivity to 10a. In the N-linked series, introduction of more than three nitrogen atoms into the heterocycle led to a reduction in receptor selectivity. Thus, tetrazole 14d has 8-fold higher affinity for 5-HT<sub>2C</sub> receptors than triazole 10a and has generally lower receptor selectivity. Removal of the methylene bridge of 10a to give the conjugated triazole 18a reduced selectivity for 5-HT<sub>1D</sub> receptors over 5-HT<sub>1A</sub> and 5-HT<sub>2C</sub>, but  $\geq$ 100-fold selectivity over 5-HT<sub>2A</sub> and 5-HT<sub>3</sub> was maintained. The conjugated 2-methylimidazole 18b, however, has improved selectivity for 5-HT<sub>1D</sub> over all the other serotonin receptors measured, compared with the methylenebridged analogue 10c. In the carbon-linked triazole series, the 4-methyl-1,2,4-triazole 26 has the optimal 5-HT<sub>1D</sub> receptor selectivity profile and is comparable to the nitrogen-linked triazole 10a. Interestingly, although the acidic N-H tetrazole 28 showed poor affinity for 5-HT<sub>1D</sub> receptors, it has high affinity and selectivity for 5-HT<sub>1A</sub> receptors. The N-1 alkyltetrazoles 35a and **36a** have higher selectivity for 5-HT<sub>1D</sub> receptors than the N-2 alkylation adducts **35b** and **36b**, with the ethyl tetrazole **36a** having the best profile. Therefore, across the triazole and tetrazole series, substitution of the azole ring  $\alpha$  to the methylene bridge always gives compounds with higher affinity and selectivity for 5-HT<sub>1D</sub> receptors, compared with the  $\beta$ -substituted products. Triazole 26 and tetrazoles 35a and 36a are more selective than analogous compounds in the previously reported 1,2,4oxadiazole series in which only substitution  $\beta$  to the methylene bridge was possible.<sup>21</sup> The binding selectivity of 10a for 5-HT<sub>1D</sub> receptors in human cortical membranes was compared with sumatriptan, and the results are shown in Table 5. Triazole 10a and sumatriptan have comparable 5-HT<sub>1D</sub> receptor selectivity in human brain cortex.

**Functional Activity**. The *in vitro* functional activity of the compounds for 5-HT<sub>1D</sub> receptors was assessed on the New Zealand white rabbit saphenous vein preparation. In this model, contractions evoked by agonists are considered to be mediated by 5-HT<sub>1</sub>-like receptors.<sup>30</sup> Agonist potencies were calculated as  $pEC_{50}$  values from plots of percentage 5-HT (1  $\mu$ M) response against concentration of the agonist. The results are shown in Table 6. Triazole 10a is a potent agonist in the preparation with potency and efficacy comparable to that of 5-HT. Compound 10a is more potent in this assay than the aminooxadiazole 4b and sumatriptan. The increased affinity observed on conjugating the heterocycle with indole, for compounds 18a,b, is reflected in the higher potency of these compounds in the functional assay. Similarly, the 1-substituted tetrazoles 35a and 36a are more potent than the 2-substituted

Table 4. Selectivity of Imidazole, Triazole, and Tetrazole Derivativies in Binding to 5-HT<sub>1D</sub> Serotonin Receptors

	$\mathrm{plC}_{50}\pm\mathrm{SEM}^a$					
compd	5- $\mathrm{HT_{1D}}^b$	5-HT <sub>1A</sub> <sup>c</sup>	$5\text{-}\mathrm{HT}_{2\mathrm{C}}{}^d$	5- $\mathrm{HT}_{2\mathrm{A}}^{e}$	5-HT <sub>3</sub> /	
sumatriptan (3)	$7.7 \pm 0.1$	$6.3 \pm 0.2$	5.1	<5.0	< 5.0	
L-695,894 (4b)	$7.6\pm0.21$	$6.5\pm0.05$	$5.9 \pm 0.09$	6.4	5.6	
MK-462 (10a)	$7.3 \pm 0.08$	$6.5\pm0.08$	$5.1 \pm 0.02$	$5.2\pm0.06$	$5.4 \pm 0.06$	
10b	$7.5\pm0.06$	$6.8\pm0.02$	$5.4 \pm 0.14$	5.5	5.9	
10c	$7.2\pm0.03$	6.5	$5.5\pm0.02$	5.1	5.8	
14 <b>a</b>	7.3	6.9	•	5.2		
14 <b>c</b>	$7.4 \pm 0.01$	$6.8\pm0.07$	$5.7 \pm 0.10$	5.3	5.9	
1 <b>4d</b>	$7.4 \pm 0.08$	6.7	$6.0 \pm 0.14$	5.7	5.5	
18 <b>a</b>	$7.7\pm0.03$	7.6	6.3	5.0	5.0	
18 <b>b</b>	$8.1\pm0.01$	7.2	5.5	<5.0	5.5	
21b	$7.0\pm0.14$	6.5	<5.0	5.4	5.0	
26	7.6	6.5	5.1	<5.0	5.1	
<b>2</b> 8	5.4	7.2	<5.0	<5.0	<5.0	
35a	7.6	$6.7\pm0.19$	$5.3\pm0.21$	<5.0	5.3 <sup>e</sup>	
36a	$7.6\pm0.16$	6.5	<5.0	<5.0	<5.0	
35b	$7.1\pm0.05$	$6.2\pm0.09$	$5.6 \pm 0.23$	5.6	5.6	
36b	6.9	6.3	5.6	5.8	5.6	

<sup>a</sup> SEM = standard error of the mean from  $n \ge 3$ . Where SEM is not quoted, the figures are the mean of two independent determinations typically with individual values within  $\pm (10-15)\%$  of the mean. <sup>b</sup> Displacement of [<sup>3</sup>H]-5-HT binding to 5-HT<sub>1D</sub> recognition sites in pig caudate membranes. <sup>c</sup> Displacement of [<sup>3</sup>H]-8-OH-DPAT from pig cortex. <sup>d</sup> Displacement of [<sup>3</sup>H]mesulergine from pig cortex. <sup>e</sup> Displacement of [<sup>3</sup>H]ODB from rat cortex homogenates. <sup>f</sup> Displacement of [<sup>3</sup>H]Q-ICS 205-930 from rat cortex homogenates. <sup>g</sup> Value derived from a single determination.

**Table 5.** Selectivity of 10a in Binding to 5-HT Receptors inHuman Brain Cortex

	$\mathrm{pIC}_{50} \pm \mathrm{SEM}^a$				
compd	$5-HT_{1D}$	5-HT <sub>1A</sub>	$5\text{-}HT_{2C}$	$5-HT_{2A}$	$5-HT_3$
10a	$7.1 \pm 0.2$	$6.4 \pm 0.07$	< 5.0	$5.1 \pm 0.2$	< 5.0
sumatriptan (a)	$1.4 \pm 0.2$	$0.4 \pm 0.04$	~5.0	$0.1 \pm 0.12$	<b>~</b> 0.0

 $^{a}$  See corresponding footnote of Table 4. The radioligands used are as shown in Table 4.

**Table 6.** In Vitro Functional Activity of Imidazole, Triazole, and Tetrazole Derivatives

compd	$\mathrm{pEC}_{50}{}^a$	relative maximum <sup>b</sup>
5-HT (1)	6.8	1.0
sumatriptan ( <b>3</b> )	6.2	1.0
4b	6.3	0.9
10a	6.6	1.0
1 <b>0b</b>	6.3	0.8
10c	6.2	0.7
1 <b>4</b> a	6.0	1.0
14 <b>c</b>	6.7	1.0
1 <b>4d</b>	6.7	0.8
18 <b>a</b>	7.2	1.2
18 <b>b</b>	6.8	1.0
21b	6.2	0.7
35a	6.6	1.0
36a	6.4	1.0
35b	6.0	1.0
36b	6.2	0.7

<sup>a</sup> Contraction of the New Zealand white rabbit saphenous vein. The figures are the mean of two independent determinations typically with individual values within  $\pm (10-15)\%$  of the mean. <sup>b</sup> Relative maximum = relative efficacy of the agonist with respect to 1  $\mu$ M 5-HT.

analogues **35b** and **36b**. A detailed description of the *in vitro* and *in vivo* functional activity of these compounds, in particular for triazole **10a**, will be presented shortly.<sup>31</sup>

**Pharmacokinetics**. Conclusions from pharmacokinetic studies in the 1,2,4-oxadiazole series were (a) all compounds containing benzyl substituents showed low oral bioavailability and small substituents on the heterocycle, e.g., methyl and amino, were best; (b) primary tryptamines gave low oral bioavailability presumably because of extensive metabolism by monoamine oxidase; and (c) measurement of brain/plasma ratios suggested



Figure 1. Concentrations of 10a benzoate and sumatriptan succinate in plasma after oral administration (3 mg kg<sup>-1</sup>) to rats.

that conjugated heterocycles show greater CNS penetration than the methylene-linked analogues, presumably because of the higher hydrophobicity of these compounds.

With these results in mind, the compounds of this study were designed to identify an optimal oral absorption profile in combination with low brain penetration. Triazole 10a was shown to have the most desired pharmacokinetic profile in rats, following oral administration (see Figure 1), with high bioavailability (76%; cf. sumatriptan, 44%) and rapid absorption ( $t_{max} = 0.75$  h). The plasma half-life ( $t_{1/2}$ ) of 10a in rat was shown to be 0.8 h (cf. sumatriptan, 1.1 h) and the steady state volume of distribution ( $V_{ss}$ ) 6.3 L/kg (cf. sumatriptan, 4.6 L/kg). After oral administration, triazole 10a could not be detected in rat brain at any time point (<20 ng/mL). Following intravenous administration (iv), the brain/plasma ratios for 10a were as follows: at 0.083 h, 0.07; at 0.25 h, 0.1; and at 0.5 h, 0.14 (cf. sumatriptan,

0.04, 0.09, and 0.1, respectively). This datum suggests that **10a** only poorly penetrates the CNS after iv dosing.<sup>32</sup> Both the C-linked triazole **26** and the 1-methyltetrazole **35a** showed poor oral absorption in rats. On the basis of its 5-HT<sub>1D</sub> receptor selectivity profile and oral bioavailability, **10a** was chosen as the clinical candidate to investigate efficacy as an antimigraine agent, and results from these studies will be published in due course.

# Conclusions

A novel series of imidazole-, triazole-, and tetrazolesubstituted indoles has been identified which are potent and selective agonists for the 5-HT<sub>1D</sub> receptor. The heterocycle can be linked to indole through either N or C, and in the N-linked series, simple unsubstituted imidazoles, triazoles, and tetrazoles have high affinity and good selectivity for the 5-HT<sub>1D</sub> receptor. The results suggest that for high affinity a H-bond acceptor interaction with a  $\beta$ -N in the azole ring is required. As reported for the oxadiazole series, directly linking the heterocycle to indole leads to a 5-10-fold increase in affinity. In a series of N-methyl-substituted triazoles and tetrazoles, linked to indole through C, optimal potency and selectivity were found when the heterocycle is substituted  $\alpha$ to the methylene bridge. This may be a consequence of either a more favorable H-bond interaction or a more positive hydrophobic interaction. The 1,2,4-triazole 10a was shown to have the optimal pharmacokinetic profile with rapid oral absorption and high bioavailability and is currently undergoing clinical trials for the treatment of migraine.

# **Experimental Section**

Chemical Methods: General Directions. Except where otherwise stated, the following procedures were adopted. <sup>1</sup>H NMR spectra were recorded at 360 MHz on a Brucker AM360 instrument and mass spectra with a VG70-250 mass spectrometer. Organic solvents were purified when necessary by the methods described by Perrin et al. (Perrin, D. D.; Armarego, W. L. F.; Perrin, D. R. Purification of Laboratory Chemicals; Pergamon: Oxford, 1966) or were purchased from Aldrich Chemical Co., Sureseal. All solutions were dried over sodium sulfate and evaporated on a Büchi rotary evaporator at reduced pressure. Thin layer chromatography and preparative chromatography were performed on silica gel, with use of plates (Merck Art. No. 5719) and columns (Merck Art. No. 7734). log D's were determined using 1-octanol and pH 7.4 buffer by the shake flask method. Microanalyses were performed by Butterworth Laboratories Ltd., Middlesex, U.K., and are within  $\pm 0.4$  unless otherwise noted. Melting points are uncorrected.

General Procedure for the Preparation of 1-(Triazolylmethyl)- and 1-(Imidazolylmethyl)-4-nitrobenzenes 7a-c. 1-(1,2,4-Triazol-1-ylmethyl)-4-nitrobenzene (7a). 4-Nitrobenzyl bromide (21.6 g, 0.1 mol) was added to a rapidly stirred suspension of 1,2,4-triazole sodium salt (9.1 g, 0.1 mol) in anhydrous DMF (100 mL) and the mixture stirred at 25 °C for 16 h. Ethyl acetate (400 mL) and water (250 mL) were added to the reaction mixture and the layers separated. The organic phase was washed with water (3×), dried, and evaporated. The crude product was chromatographed on silica gel, eluting with ethyl acetate, to give 7a (10.6 g, 52%): mp 101-102 °C; <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  5.47 (2H, s, CH<sub>2</sub>), 7.40 (2H, d, J = 9.0 Hz, Ar-H), 8.02 and 8.18 (each 1H, each s, triazole-H), 8.23 (2H, d, J = 9.0 Hz, Ar-H). Anal. (C<sub>9</sub>H<sub>8</sub>N<sub>4</sub>O<sub>2</sub>) H, N; C: calcd, 52.9; found, 53.4.

**7b:** 18%; mp 55–56 °C; <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  5.26 (2H, s, CH<sub>2</sub>), 6.92 (1H, s, imidazole-H), 7.16 (1H, s, imidazole-H), 7.28 (2H,

d, J = 9.5 Hz, Ar-H), 7.62 (1H, s, imidazole-H), 8.22 (2H, d, J = 9.5 Hz, Ar-H).

**7c:** 10.5%; <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  2.34 (3H, s, CH<sub>3</sub>), 5.16 (2H, s, CH<sub>2</sub>), 6.67 (1H, d, J = 1.3 Hz, imidazole-H), 7.03 (1H, d, J = 1.3 Hz, imidazole-H), 7.19 (2H, d, J = 9.5 Hz, Ar-H), 8.22 (2H, d, J = 9.5 Hz, Ar-H).

1-(1,2,3-Triazol-1-ylmethyl)-4-nitrobenzene (11a) and 1-(1,2,3-Triazol-2-ylmethyl)-4-nitrobenzene (11b). 4-Nitrobenzyl bromide (25.4 g, 0.12 mol) was added to a solution of 1*H*-1,2,3-triazole (8.12 g, 0.12 mol) and triethylamine (11.88 g, 0.12 mol) in anhydrous acetonitrile (150 mL). The mixture was refluxed for 1 h and then cooled to room temperature and the precipitated triethylamine hydrobromide filtered off. The solvent was evaporated and the resultant crude product chromatographed on silica gel eluting with CH<sub>2</sub>Cl<sub>2</sub>/MeOH (100:0  $\rightarrow$  95:5) to give 11a (13 g, 54%) and 11b (2.25 g, 9%). 11a was isolated as the more polar isomer: mp 110-111 °C; <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  5.72 (2H, s, CH<sub>2</sub>), 7.38 (2H, d, J = 9.0 Hz, Ar-H), 7.64 and 7.78 (each 1H, each s, triazole-H), 8.18 (2H, d, J = 9.0 Hz, Ar-H). Anal. (C<sub>3</sub>H<sub>8</sub>N<sub>4</sub>O<sub>2</sub>) C, H, N.

11b: mp 110–112 °C; <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  5.72 (2H, s, CH<sub>2</sub>), 7.40 (2H, d, J = 9.0 Hz, Ar-H), 7.66 (2H, s, triazole-H), 8.18 (2H, d, J = 9.0 Hz, Ar-H). Anal. (C<sub>9</sub>H<sub>8</sub>N<sub>4</sub>O<sub>2</sub>) C, H, N.

1-(Tetrazol-1-ylmethyl)-4-nitrobenzene (11c) and 1-(Tetrazol-2-ylmethyl)-4-nitrobenzene (11d). The title compounds were prepared from 4-nitrobenzyl bromide and 1*H*tetrazole using the procedure described for 11a,b. The more polar, major product was identified as 11c (75%): mp 93 °C; <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  5.73 (2H, s, CH<sub>2</sub>), 7.46 and 8.27 (each 2H, each d, J = 8.7 Hz, Ar-H), 8.64 (1H, s, tetrazole-H). Anal. (C<sub>8</sub>H<sub>7</sub>N<sub>5</sub>O<sub>2</sub>) C, H; N: calcd, 34.1; found, 34.6.

**11d:** 17%; mp 127–128 °C; <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  5.92 (2H, s, CH<sub>2</sub>), 7.53 and 8.25 (each 2H, each d, J = 8.7 Hz, Ar-H), 8.56 (1H, s, tetrazole-H). Anal. (C<sub>8</sub>H<sub>7</sub>N<sub>5</sub>O<sub>2</sub>) C, H, N.

General Procedure for the Preparation of N,N-Dimethyltryptamines 10a-c and 14a-d. N,N-Dimethyl-2-[5-(1,2,4-triazol-1-ylmethyl)-1H-indol-3-yl]ethylamine Benzoate (10a). A solution of 7a (10.0 g, 49.0 mmol) in EtOH (50 mL), ethyl acetate (50 mL), and 2 N HCl (20 mL) was hydrogenated over 10% Pd-C (1.0 g) in a Parr shake apparatus at 40 psi for 0.2 h. The catalyst was removed by filtration through Hyflo, the solvent evaporated, and the residue azeotroped with EtOH (3×) to give **8a** (10.6 g, 100%) which was characterized as the free base: mp 126-128 °C; <sup>1</sup>H NMR (CD<sub>3</sub>OD)  $\delta$  5.23 (2H, s, CH<sub>2</sub>), 6.68 and 7.08 (each 2H, each d, J = 9.0 Hz, Ar-H), 7.95 and 8.39 (each 1H, each s, triazole-H). Anal. (C<sub>9</sub>H<sub>10</sub>N<sub>4</sub>) C, H, N.

A solution of NaNO<sub>2</sub> (3.28 g, 48.0 mmol) in H<sub>2</sub>O (20 mL) was added to a cooled (-10 °C) solution of **8a** (10.0 g, 48.0 mmol) in concentrated HCl (40 mL), at a such a rate that the temperature did not exceed 0 °C. The mixture was stirred at 0 °C for 0.1 h and then added portionwise to a cooled (-10 °C) and rapidly stirred solution of SnCl<sub>2</sub>·2H<sub>2</sub>O (40.0 g, 0.18 mol) in concentrated HCl (40 mL), at such a rate that the temperature did not exceed -5 °C. The solution was warmed to room temperature, basified with 20% aqueous NaOH solution, and extracted with ethyl acetate (3×). The combined extracts were dried and evaporated to give **9a** (5.0 g, 56%): mp 109-112 °C; <sup>1</sup>H NMR (DMSO-d<sub>6</sub>)  $\delta$  3.93 (2H, br s, NH<sub>2</sub>), 5.20 (2H, s, CH<sub>2</sub>), 6.73 and 7.08 (each 2H, each d, J = 8.0 Hz, Ar-H), 7.92 and 8.57 (each 1H, each s, triazole-H).

To a solution of **9a** (5.0 g, 26.4 mmol) in EtOH/H<sub>2</sub>O (5:1, 180 mL) and 5 N HCl (4.5 mL) was added 4-chlorobutanal dimethyl acetal<sup>26</sup> (3.22 g, 21.1 mmol), and the solution was refluxed for 4 h. The solvents were evaporated and the residue chromatographed on silica gel, eluting with CH<sub>2</sub>Cl<sub>2</sub>/EtOH/NH<sub>3</sub> (30:8:1), to give 2-[5-(1,2,4-triazol-1-ylmethyl)-1H-indol-3-yl]-ethylamine (2.4 g, 38%). A solution of formaldehyde (0.30 g of a 37%, w/v, solution, 3.7 mmol) in MeOH (10 mL) was added to a stirred mixture of 2-[5-(1,2,4-triazol-1-ylmethyl)-1H-indol-3-yl]ethylamine (0.36 g, 1.5 mmol), NaCNBH<sub>3</sub> (0.225 g, 3.6 mmol), and glacial acetic acid (0.45 g, 7.5 mmol), in MeOH (10 mL), at 0 °C. The solution was warmed to 25 °C and stirred for 2 h before adding saturated K<sub>2</sub>CO<sub>3</sub> solution (5 mL). The MeOH was removed under vacuum, H<sub>2</sub>O (10 mL) was added, and the mixture was extracted with ethyl acetate (3×).

### Potent Agonists for 5-HT<sub>1D</sub> Receptors

The combined extracts were washed with brine  $(1\times)$  and dried and the solvent evaporated. The crude product was chromatographed on silica gel eluting with CH<sub>2</sub>Cl<sub>2</sub>/EtOH/NH<sub>3</sub> (40:8:1) to give 10a (0.21 g, 52%). The benzoate salt was prepared. 10a: mp 178–180 °C; MS m/z 269 (M<sup>+</sup>); <sup>1</sup>H NMR (D<sub>2</sub>O)  $\delta$  2.91 (6H, s, N(CH<sub>3</sub>)<sub>2</sub>), 3.19 and 3.43 (each 2H, each t, J = 7.0 Hz, 1-CH<sub>2</sub> and 2-CH<sub>2</sub>), 5.49 (2H, s, CH<sub>2</sub>), 7.20 (1H, dd, J = 1.6and 8.4 Hz, 6'-CH), 7.34 (1H, s, 2'-CH), 7.52 (1H, d, J = 8.4Hz, 7'-CH), 7.43–7.53 (3H, m, benzoic acid-H), 7.62 (1H, d, J = 1.6 Hz, 4'-CH), 7.87–7.90 (2H, m, benzoic acid-H), 8.05 (1H, s, triazole-H), 8.52 (1H, s, triazole-H). Anal. (Cl<sub>15</sub>H<sub>19</sub>N<sub>5</sub>·C<sub>6</sub>H<sub>5</sub>-COOH) C, H, N.

The following N,N-dimethyltryptamines were prepared using the general procedure.

**N,N-Dimethyl-2-[5-(imidazol-1-ylmethyl)-1H-indol-3-yl]ethylamine dioxalate** (10b): mp 165-166 °C (MeOH/ Et<sub>2</sub>O); <sup>1</sup>H NMR (D<sub>2</sub>O)  $\delta$  2.92 (6H, s, N(CH<sub>3</sub>)<sub>2</sub>), 3.24 and 3.48 (each 2H, each t, J = 7.7 Hz, 1-CH<sub>2</sub> and 2-CH<sub>2</sub>), 5.50 (2H, s, CH<sub>2</sub>), 7.27 (1H, dd, J = 1.5 and 8.4 Hz, 6'-CH), 7.37 (1H, s, 2'-CH), 7.45 and 7.49 (each 1H, each s, imidazole-H), 7.56 (1H, d, J = 8.4 Hz, 7'-CH), 7.75 (1H, s, 4'-CH), 8.78 (1H, s, imidazole-H). Anal. (C<sub>16</sub>H<sub>20</sub>N<sub>4</sub>•2.05(COOH)<sub>2</sub>) C, H, N.

*N*,*N*-Dimethyl-2-[5-[(2-methylimidazol-1-yl)methyl]-1*H*-indol-3-yl]ethylamine trioxalate (10c): mp 160−163 °C (MeOH/Et<sub>2</sub>O); MS m/z 282 (M<sup>+</sup>); <sup>1</sup>H NMR (D<sub>2</sub>O)  $\delta$  2.65 (3H, s, CH<sub>3</sub>), 2.92 (6H, s, N(CH<sub>3</sub>)<sub>2</sub>), 3.25 and 3.50 (each 2H, each t, *J* = 7.3 Hz, 1-CH<sub>2</sub> and 2-CH<sub>2</sub>), 5.42 (2H, s, CH<sub>2</sub>), 7.18 (1H, d, *J* = 8.4 Hz, 6'-CH), 7.31−7.40 (3H, m, imidazole-H and 2'-CH), 7.56 (1H, d, *J* = 8.4 Hz, 7'-CH), 7.66 (1H, s, 4'-CH). Anal. (C<sub>17</sub>H<sub>22</sub>N<sub>4</sub>·2.8(COOH)<sub>2</sub>) C, H, N.

**N,N-Dimethyl-2-[5-(1,2,3-triazol-1-ylmethyl)-1H-indol-3-yl]ethylamine hydrogen oxalate** (14a): mp 210–212 °C; <sup>1</sup>H NMR (D<sub>2</sub>O)  $\delta$  2.90 (6H, s, N(CH<sub>3</sub>)<sub>2</sub>), 3.22 and 3.46 (each 2H, each t, J = 7.4 Hz, 1-CH<sub>2</sub> and 2-CH<sub>2</sub>), 5.72 (2H, s, CH<sub>2</sub>), 7.24 (1H, dd, J = 1.6 and 8.4 Hz, 6'-CH), 7.36 (1H, s, 2'-CH), 7.52 (1H, d, J = 8.4 Hz, 7'-CH), 7.66 and 7.79 (each 1H, each s, triazole-H), 8.00 (1H, d, J = 1.6 Hz, 4'-CH). Anal. (C<sub>15</sub>H<sub>19</sub>N<sub>5</sub>·1.1(COOH)<sub>2</sub>·0.15H<sub>2</sub>O) C, H, N.

N,N-Dimethyl-2-[5-(1,2,3-triazol-2-ylmethyl)-1*H*-indol-3-yl]ethylamine hydrogen oxalate (14b): mp 204–205 °C; <sup>1</sup>H NMR (D<sub>2</sub>O)  $\delta$  2.90 (6H, s, N(CH<sub>3</sub>)<sub>2</sub>), 3.21 and 3.46 (each 2H, each t, J = 7.4 Hz, 1-CH<sub>2</sub> and 2-CH<sub>2</sub>), 5.74 (2H, s, CH<sub>2</sub>), 7.22 (1H, dd, J = 1.6 and 8.4 Hz, 6'-CH), 7.34 (1H, s, 2'-CH), 7.50 (1H, d, J = 8.4 Hz, 7'-CH), 7.64 (1H, s, 4'-CH), 7.78 (2H, s, triazole-H). Anal. (C<sub>15</sub>H<sub>19</sub>N<sub>5</sub>(COOH)<sub>2</sub>·0.25H<sub>2</sub>O) C, H, N.

**N,N-Dimethyl-2-[5-(tetrazol-1-ylmethyl)-1H-indol-3-yl]ethylamine succinate** (14c): mp 55–56 °C (hygroscopic); <sup>1</sup>H NMR (D<sub>2</sub>O)  $\delta$  2.93 (6H, s, N(CH<sub>3</sub>)<sub>2</sub>), 3.23 and 3.48 (each 2H, each t, J = 7.5 Hz, 1-CH<sub>2</sub> and 2-CH<sub>2</sub>), 5.81 (2H, s, CH<sub>2</sub>), 7.28 (1H, dd, J = 1.7 and 8.4 Hz, 6'-CH), 7.39 (1H, s, 2'-CH), 7.56 (1H, d, J = 8.4 Hz, 7'-CH), 7.75 (1H, s, 4'-CH), 9.20 (1H, s, tetrazole-H). Anal. (C<sub>14</sub>H<sub>18</sub>N<sub>6</sub>•0.75(CH<sub>2</sub>COOH)<sub>2</sub>) C, H, N.

**N,N-Dimethyl-2-[5-(tetrazol-2-ylmethyl)-1H-indol-3-yl]-ethylamine hydrogen oxalate** (14d): mp 198–199 °C (EtOH/Et<sub>2</sub>O); <sup>1</sup>H NMR (D<sub>2</sub>O)  $\delta$  2.91 (6H, s, N(CH<sub>3</sub>)<sub>2</sub>), 3.23 and 3.48 (each 2H, each t, J = 7.4 Hz, 1-CH<sub>2</sub> and 2-CH<sub>2</sub>), 6.01 (2H, s, CH<sub>2</sub>), 7.30 (1H, dd, J = 1.6 and 8.4 Hz, 6'-CH), 7.37 (1H, s, 2'-CH), 7.53 (1H, d, J = 8.4 Hz, 7'-CH), 7.76 (1H, s, 4'-CH), 8.74 (1H, s, tetrazole-H). Anal. (C<sub>14</sub>H<sub>18</sub>N<sub>6</sub>·(COOH)<sub>2</sub>·0.2C<sub>2</sub>H<sub>5</sub>-OH) C, H, N.

Alternative Procedure for the Preparation of 10a. To a solution of the hydrochloride salt of **9a** (5.0 g, 22.2 mmol) in 4% H<sub>2</sub>SO<sub>4</sub> (190 mL) was added 4-(*N*,*N*-dimethylamino)butanal dimethyl acetal<sup>28</sup> (5.35 g, 33.2 mmol), and the solution was refluxed for 2 h. The mixture was cooled to room temperature, ethyl acetate (400 mL) was added, and the aqueous was basified with 5 N NaOH solution. The aqueous was separated and extracted with ethyl acetate (2×), and the combined extracts were dried and evaporated. The crude product was chromatographed on silica gel eluting with CH<sub>2</sub>Cl<sub>2</sub>/EtOH/NH<sub>3</sub> (40:8:1) to give 10a (2.2 g, 37%).

1-(1,2,4-Triazol-1-yl)-4-nitrobenzene (15a). 1-Fluoro-4nitrobenzene (25.0 g, 0.18 mol) was added to a rapidly stirred suspension of 1,2,4-triazole sodium salt (17.7 g, 0.19 mol) in anhydrous DMF (150 mL) and the mixture stirred at room temperature for 16 h. Ethyl acetate (500 mL) and water (300 mL) were added to the reaction mixture which was then stirred for 0.1 h and the layers separated. The organic phase was washed with water (3×) and dried and the solvent evaporated to give 15a (24.8 g, 74%): mp 197–198 °C; MS *m*/*z* 190 (M<sup>+</sup>); <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  7.92 (2H, d, *J* = 9.1 Hz, Ar-H), 8.17 (1H, s, triazole-H), 8.40 (2H, d, *J* = 9.1 Hz, Ar-H), 8.48 (1H, s, triazole-H). Anal. (C<sub>8</sub>H<sub>6</sub>N<sub>4</sub>O<sub>2</sub>) H, N; C: calcd, 50.5; found, 51.2.

1-(2-Methylimidazol-1-yl)-4-nitrobenzene (15b). Sodium hydride (60% dispersion in oil, 4.87 g, 0.122 mol) was added to a solution of 2-methylimidazole (10.0 g, 0.122 mol) in DMF (100 mL) and the mixture stirred for 0.1 h. 1-Fluoro-4-nitrobenzene (17.18 g, 0.122 mol) was added and the mixture stirred at room temperature for 16 h. Water (150 mL) and ethyl acetate (250 mL) were added to the mixture, which was stirred for 0.1 h, and the layers separated. The aqueous phase was extracted with ethyl acetate (3×), and the combined extracts were washed with H<sub>2</sub>O (3×), dried, and evaporated to give 15b (11.5 g, 47%): mp 139-140 °C; <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$ 2.24 (3H, s, CH<sub>3</sub>), 7.06 and 7.10 (each 1H, each d, J = 1.5 Hz, imidazole-H), 7.50 and 8.38 (each 2H, each d, J = 9.5 Hz, Ar-H). Anal. (C<sub>10</sub>H<sub>9</sub>N<sub>3</sub>O<sub>2</sub>) C, H, N.

**N,N-Dimethyl-2-[5-(1,2,4-triazol-1-yl)-1H-indol-3-yl]eth-ylamine Dioxalate (18a).** The title compound was prepared from 15a using the general procedure described for 10a: mp 210 °C (MeOH/Et<sub>2</sub>O); <sup>1</sup>H NMR (D<sub>2</sub>O)  $\delta$  2.92 (6H, s, N(CH<sub>3</sub>)<sub>2</sub>), 3.25 and 3.50 (each 2H, each t, J = 7.4 Hz, 1-CH<sub>2</sub> and 2-CH<sub>2</sub>), 7.44 (1H, s, 2'-CH), 7.47 (1H, dd, J = 2.0 and 8.7 Hz, 6'-CH), 7.63 (1H, d, J = 8.7 Hz, 7'-CH), 7.88 (1H, d, J = 2.0 Hz, 4'-CH), 8.36 and 9.05 (each 1H, each s, triazole-H). Anal. (C<sub>14</sub>H<sub>17</sub>N<sub>5</sub>·1.9(COOH)<sub>2</sub>) C, H, N.

*N,N*-Dimethyl-2-[5-(2-methylimidazol-1-yl)-1*H*-indol-3yl]ethylamine Sesquioxalate (18b). The title compound was prepared from 15b using the general procedures: mp 185–186 °C; <sup>1</sup>H NMR (D<sub>2</sub>O)  $\delta$  2.55 (3H, s, CH<sub>3</sub>), 2.93 (6H, s, N(CH<sub>3</sub>)<sub>2</sub>), 3.26 and 3.51 (each 2H, each t, J = 7.4 Hz, 1-CH<sub>2</sub> and 2-CH<sub>2</sub>), 7.30 (1H, dd, J = 2.0 and 8.7 Hz, 6'-CH), 7.48 (1H, d, J = 2.1 Hz, imidazole-H), 7.51–7.53 (2H, m, imidazole-H and 2'-CH), 7.70 (1H, d, J = 8.7 Hz, 7'-CH), 7.79 (1H, d, J = 2.0 Hz, 4'-CH). Anal. (C<sub>16</sub>H<sub>20</sub>N<sub>4</sub>•1.55(COOH)<sub>2</sub>•0.1EtOH) C, H, N.

*N,N*-Dimethyl-2-[5-[(1-methyl-1,2,4-triazol-3-yl)methyl]-1*H*-indol-3-yl]ethylamine Trihydrochloride (21a) and *N,N*-Dimethyl-2-[5-[(1-methyl-1,2,4-triazol-5-yl)methyl]-1*H*-indol-3-yl]ethylamine Dioxalate (21b). A solution of *N,N*-dimethyl-2-[5-(cyanomethyl)-1*H*-indol-3-yl]ethylamine<sup>21</sup> (19) (5.0 g, 22.0 mmol) in EtOH (50 mL) was saturated with HCl (gas) and stirred at room temperature for 16 h. The solvent was evaporated under reduced pressure to give 20 (6.0 g, 92%): <sup>1</sup>H NMR (DMSO- $d_6$ )  $\delta$  1.29 (3H, t, J = 7.0 Hz, CH<sub>3</sub>), 2.83 (6H, s, N(CH<sub>3</sub>)<sub>2</sub>), 3.13 and 3.31 (each 2H, each t, J = 7.5 Hz, 1-CH<sub>2</sub> and 2-CH<sub>2</sub>), 4.04 (2H, s, CH<sub>2</sub>), 4.42 (2H, q, J = 7.0 Hz, *CH*<sub>2</sub>CH<sub>3</sub>), 7.08 (1H, dd, J = 1.5 and 8.4 Hz, 6'-CH), 7.27 (1H, s, 2'-CH), 7.37 (1H, d, J = 8.4 Hz, 7'-CH), 7.48 (1H, br s, NH), 7.71 (1H, s, 4'-CH).

A mixture of 20 (3.0 g, 10.15 mmol), methylhydrazine (0.69 g, 15.0 mmol), and triethylamine (2.6 g, 25.4 mmol) in EtOH (30 mL) was stirred at room temperature for 3 h. The solvent was evaporated and the product dissolved in formic acid (98%, 3.3 mL). The solution was stirred at room temperature for 0.5 h and then at reflux for 2 h. After cooling to room temperature, the mixture was poured into K<sub>2</sub>CO<sub>3</sub> solution (75 mL) and extracted with ethyl acetate  $(4 \times)$ . The combined extracts were dried, the solvent was evaporated, and the crude product was chromatographed on silica gel eluting with CH2-Cl<sub>2</sub>/EtOH/NH<sub>3</sub> (40:8:1) to give two products. The less polar isomer was identified as  $\mathbf{21b}$  (0.36 g, 12.5%) and the dioxalate salt was prepared. 21b: mp 135–137 °C; <sup>1</sup>H NMR (D<sub>2</sub>O)  $\delta$ 2.91 (6H, s, N(CH<sub>3</sub>)<sub>2</sub>), 3.23 and 3.48 (each 2H, each t, J = 7.3Hz, 1-CH<sub>2</sub> and 2-CH<sub>2</sub>), 3.95 (3H, s, CH<sub>3</sub>), 4.48 (2H, s, CH<sub>2</sub>), 7.13 (1H, dd, J = 1.5 and 8.4 Hz, 6'-CH), 7.37 (1H, s, 2'-CH), 7.53 (1H, d, J = 8.4 Hz, 7'-CH), 7.57 (1H, s, 4'-CH), 8.32 (1H, s, triazole-H). Anal.  $(C_{16}H_{21}N_5 \cdot 2.25(COOH)_2)$  C, H; N: calcd, 14.4; found, 13.9.

The more polar product was identified as 21a (0.18 g, 6.3%). Treatment of 21a with ethereal HCl gave the trihydrochloride salt: mp <40 °C (hygroscopic); <sup>1</sup>H NMR (D<sub>2</sub>O)  $\delta$  2.91 (6H, s, N(CH<sub>3</sub>)<sub>2</sub>), 3.23 and 3.49 (each 2H, each t, J = 7.4 Hz, 1-CH<sub>2</sub> and 2-CH<sub>2</sub>), 3.95 (3H, s, CH<sub>3</sub>), 4.27 (2H, s, CH<sub>2</sub>), 7.17 (1H, dd, J = 1.5 and 8.5 Hz, 6'-CH), 7.34 (1H, s, 2'-CH), 7.50 (1H, d, J = 8.5 Hz, 7'-CH), 7.60 (1H, s, 4'-CH), 8.88 (1H, s, triazole-H). Anal. (C<sub>16</sub>H<sub>21</sub>N<sub>5</sub>·3HCl·0.35Et<sub>2</sub>O) C, H, N.

1-[(4-Methyl-1,2,4-triazol-3-yl)methyl]-4-nitrobenzene (24). A mixture of 22 (10.0 g, 43.0 mmol) and formylhydrazine (2.6 g, 43.0 mmol) in anhydrous MeOH (175 mL) was stirred at room temperature for 3 days. The solvent was removed under vacuum, the residue triturated with ether, and the precipitate filtered off. The filtrate was concentrated under reduced pressure and the resultant crude product chromatographed on silica gel eluting with  $CH_2Cl_2/MeOH$  (9:1) to give  ${\bf 23} \ (2.25 \ g, \ 22\%).$  Methylamine  $(gas) \ was \ bubbled \ through \ a$ solution of 23 (1.25 g, 5.3 mmol) in anhydrous MeOH (30 mL), for 0.25 h, and the mixture then stirred for 72 h at room temperature. The solvent was evaporated under reduced pressure and the residue chromatographed on silica gel eluting with  $CH_2Cl_2/MeOH$  (9:1) to give 24 (1.0 g, 87%) as a white crystalline solid: mp 168–170 °C; MS m/z 218 (M<sup>+</sup>); <sup>1</sup>H NMR (CDCl<sub>3</sub>) & 3.51 (3H, s, CH<sub>3</sub>), 4.31 (2H, s, CH<sub>2</sub>), 7.42 (2H, d, J = 8.7 Hz, Ar-H), 8.11 (1H, s, triazole-H), 8.20 (2H, d, J = 8.7 Hz, Ar-H). Anal.  $(C_{10}H_{10}N_4O_2)$  C, H, N.

*N,N*-Dimethyl-2-[5-[(4-methyl-1,2,4-triazol-3-yl)methyl]-1*H*-indol-3-yl]ethylamine Hydrogen Oxalate Hemihydrate (26). The title compound was prepared from 24 using the procedures described for 10a. The hydrogen oxalate hemihydrate salt was prepared. 26: mp <40 °C (hygroscopic); MS m/z 283 (M<sup>+</sup>); <sup>1</sup>H NMR (D<sub>2</sub>O)  $\delta$  2.89 (6H, s, N(CH<sub>3</sub>)<sub>2</sub>), 3.20 and 3.45 (each 2H, each t, J = 7.4 Hz, 1-CH<sub>2</sub> and 2-CH<sub>2</sub>), 3.64 (3H, s, CH<sub>3</sub>), 4.37 (2H, s, CH<sub>2</sub>), 7.12 (1H, dd, J = 1.5 and 8.4 Hz, 6'-CH), 7.34 (1H, s, 2'-CH), 7.50 (1H, d, J = 8.4 Hz, 7'-CH), 7.52 (1H, s, 4'-CH), 8.55 (1H, s, triazole-H). Anal. (C<sub>16</sub>H<sub>21</sub>N<sub>5</sub>·1.2(COOH)<sub>2</sub>·0.5H<sub>2</sub>O) C, H, N.

N,N-Dimethyl-2-[5-(tetrazol-5-ylmethyl)-1H-indol-3-yl]ethylamine Hydrochloride (28). A solution of the hydrochloride salt of 19 (0.58 g, 2.2 mmol), sodium azide (0.43 g, 6.6 mmol) and triethylamine hydrochloride (0.45 g, 3.3 mmol), in N-methylpyrrolidin-2-one (10 mL) was heated at 160 °C for 8 h; 5 N HCl (2 mL) was added, and the solvents were removed by distillation under high vacuum. Inorganic salts were removed by addition of MeOH (20 mL) and ether (30 mL). The filtrate was concentrated and the crude product chromatographed on silica gel eluting with  $EtOH/Et_2O/H_2O/NH_3$  (20: 40.8:1) to give **28** (0.45 g, 76%). The hydrochloride salt was prepared. 28: mp 223-225 °C (MeOH/Et<sub>2</sub>O); <sup>1</sup>H NMR (D<sub>2</sub>O)  $\delta$  2.90 (6H, s, N(CH<sub>3</sub>)<sub>2</sub>), 3.20 and 3.45 (each 2H, each t, J =7.4 Hz, 1-CH<sub>2</sub> and 2-CH<sub>2</sub>), 4.43 (2H, s, CH<sub>2</sub>), 7.18 (1H, dd, J =1.5 and 8.4 Hz, 6'-CH), 7.33 (1H, s, 2'-CH), 7.49 (1H, d, J = 8.4 Hz, 7'-CH), 7.56 (1H, s, 4'-CH). Anal. (C14H18N6·HCl· 0.1H<sub>2</sub>O) C, H, N.

N-(tert-Butyloxycarbonyl)-2-[5-(tetrazol-5-ylmethyl)-1H-indol-3-yl]ethylamine (30). A solution of 2-[5-(cyanomethyl)-1H-indol-3-yl]ethylamine hydrochloride (27) (2.5 g, 10.6 mmol), triethylamine hydrochloride (2.2 g, 16.0 mmol), and sodium azide (2.1 g, 32.3 mmol) in N-methylpyrrolidin-2-one (30 mL) was heated at 140 °C for 8 h; 5 N HCl (3 mL) was added, and the solvents were removed by distillation under vacuum. The residue was chromatographed on silica gel eluting with EtOH/Et<sub>2</sub>O/H<sub>2</sub>O/NH<sub>3</sub> (20:30:8:1) to give 29 (1.76 g, 69%). Triethylamine (1.5 g, 14.9 mmol) and di-tertbutyl dicarbonate (1.9 g, 7.3 mmol) were added to a stirred suspension of  $\mathbf{29}$  (1.76 g, 7.27 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (40 mL), and the mixture was stirred for 16 h at room temperature. The solvent was removed under vacuum and the crude product chromatographed on silica gel eluting with  $EtOH/Et_2O/H_2O/$  $NH_3$  (20:60:8:1) to give **30** (1.6 g, 64%): mp 95–98 °C; <sup>1</sup>H NMR  $(CD_3OD) \delta 1.41 (9H, s, (CH_3)_3), 2.87 and 3.30 (each 2H, each$ t, J = 7.4 Hz, 1-CH<sub>2</sub> and 2-CH<sub>2</sub>), 4.32 (2H, s, CH<sub>2</sub>), 6.99 (1H, d, J = 8.3 Hz, 6'-CH), 7.04 (1H, s, 2'-CH), 7.26 (1H, d, J = 8.3Hz, 7'-CH), 7.49 (1H, s, 4'-CH). Anal. (C17H22N6O2.0.8H2O) C, H, N.

*N*-(*tert*-Butyloxycarbonyl)-2-[5-[(1-methyltetrazol-5yl)methyl]-1*H*-indol-3-yl]ethylamine (31a) and *N*-(*tert*-Butyloxycarbonyl)-2-[5-[(2-methyltetrazol-5-yl)methyl]-1*H*-indol-3-yl]ethylamine (31b). Methyl iodide (2.49 g, 17.5 mmol) was added to a stirred solution of **30** (1.0 g, 2.93 mmol) and triethylamine (0.59 g, 5.85 mmol), in acetonitrile (50 mL), and the mixture stirred at room temperature for 16 h. The solvent was evaporated under reduced pressure and the residue dissolved in CH<sub>2</sub>Cl<sub>2</sub> (100 mL), washed with H<sub>2</sub>O (2×), dried, and evaporated. The crude product was chromatographed on silica gel eluting with MeOH/CH<sub>2</sub>Cl<sub>2</sub> (5:95) to give two separated products. The more polar major isomer was identified as the N-1 methylation product **31a** (0.20 g, 19%): mp 65 °C; <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  1.43 (9H, s, (CH<sub>3</sub>)<sub>3</sub>), 2.90 (2H, t, J = 6.9 Hz, 2-CH<sub>2</sub>), 3.42 (2H, br s, 1-CH<sub>2</sub>), 3.83 (3H, s, CH<sub>3</sub>), 4.40 (2H, s, CH<sub>2</sub>), 6.98 (1H, dd, J = 1.4 and 8.2 Hz, 6'-CH), 7.05 (1H, s, 2'-CH), 7.31 (1H, d, J = 8.2 Hz, 7'-CH), 7.43 (1H, s, 4'-CH), 8.18 (1H, br s, NH). Anal. (C<sub>18</sub>H<sub>24</sub>N<sub>6</sub>O<sub>2</sub>) C, H, N.

The less polar isomer was identified as the N-2 methylation product **31b** (0.10 g 10%): mp 65 °C; <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  1.43 (9H, s, (CH<sub>3</sub>)<sub>8</sub>), 2.88 (2H, t, J = 6.8 Hz, 2-CH<sub>2</sub>), 3.38–3.46 (2H, m, 1-CH<sub>2</sub>), 4.25 (3H, s, CH<sub>3</sub>), 4.32 (2H, s, CH<sub>2</sub>), 6.91 (1H, s, 2'-CH), 7.13 (1H, dd, J = 1.6 and 8.3 Hz, 6'-CH), 7.26 (1H, d, J = 8.3 Hz, 7'-CH), 7.52 (1H, s, 4'-CH), 8.43 (1H, br s, NH). Anal. (C<sub>18</sub>H<sub>24</sub>N<sub>6</sub>O<sub>2</sub>·0.2H<sub>2</sub>O) C, H; N: calcd, 23.3; found, 22.6.

N-(tert-Butyloxycarbonyl)-2-[5-[(1-ethyltetrazol-5-yl)methyl]-1H-indol-3-yl]ethylamine (33a) and N-(tert-Butyloxycarbonyl)-2-[5-[(2-ethyltetrazol-5-yl)methyl]-1Hindol-3-yl]ethylamine (33b). Ethyl iodide (7.64 mL, 95.5 mmol) was added to a stirred solution of 30 (5.74 g, 16.8 mmol) and triethylamine (4.67 mL, 33.5 mmol), in acetonitrile (220 mL), and the mixture stirred at room temperature for 16 h. The solvent was evaporated under reduced pressure, CH<sub>2</sub>Cl<sub>2</sub> (150 mL) added, and the mixture washed with  $H_2O(2\times)$ . The crude product obtained was chromatographed on silica gel eluting with Et<sub>2</sub>O/hexane (80:20)  $\rightarrow$  Et<sub>2</sub>O (100%)  $\rightarrow$  EtOAc (100%) to give two products. The more polar isomer was identified as the N-1 alkylation product 33a (2.7 g, 43%): mp 52–54 °C; <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  1.31 (3H, t, J = 7.3 Hz, CH<sub>3</sub>CH<sub>2</sub>), 1.43 (9H, s,  $(CH_3)_3$ ), 2.90 (2H, t, J = 7.0 Hz, 2-CH<sub>2</sub>), 3.41 (2H, br s, 1-CH<sub>2</sub>), 4.17 (2H, q, J = 7.3 Hz, CH<sub>2</sub>CH<sub>3</sub>), 4.39 (2H, s, CH<sub>2</sub>), 6.98 (1H, dd, J = 1.4 and 8.3 Hz, 6'-CH), 7.05 (1H, s, 2'-CH), 7.30 (1H, d, J = 8.3 Hz, 7'-CH), 7.42 (1H, s, 4'-CH), 8.18 (1H, br s, NH). Anal. (C<sub>19</sub>H<sub>26</sub>N<sub>6</sub>O<sub>2</sub>) H, N; C: calcd, 61.6; found. 62.2.

The less polar isomer was identified as the N-2 alkylation product **33b** (2.27 g, 37%): mp 108–110 °C; <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  1.43 (9H, s, (CH<sub>3</sub>)<sub>3</sub>), 1.59 (3H, t, J = 7.3 Hz, CH<sub>3</sub>CH<sub>2</sub>), 2.92 (2H, t, J = 6.8 Hz, 2-CH<sub>2</sub>), 3.38–3.48 (2H, m, 1-CH<sub>2</sub>), 4.33 (2H, s, CH<sub>2</sub>), 4.58 (2H, q, J = 6.8 Hz, CH<sub>2</sub>CH<sub>3</sub>), 7.00 (1H, s, 2'-CH), 7.18 (1H, dd, J = 1.4 and 8.3 Hz, 6'-CH), 7.29 (1H, d, J = 8.3 Hz, 7'-CH), 7.54 (1H, s, 4'-CH), 8.04 (1H, br s, NH). Anal. (C<sub>19</sub>H<sub>28</sub>N<sub>6</sub>O<sub>2</sub>) C, H, N.

General Procedure for the Preparation of Methyltetrazoles 35a,b and Ethyltetrazoles 36a,b. N,N-Dimethyl-2-[5-[(2-ethyltetrazol-5-yl)methyl]-1H-indol-3-yl]ethylamine Hydrogen Oxalate (36b). Trifluoroacetic acid (1.5 mL) was added dropwise to a stirred solution of **33b** (0.164 g, 0.44 mmol) in  $CH_2Cl_2$  (5 mL) at room temperature. The mixture was stirred for 0.75 h, and the solvents were then evaporated under reduced pressure. The residue was chromatographed on silica gel eluting with CH<sub>2</sub>Cl<sub>2</sub>/EtOH/NH<sub>3</sub> (40: 8:1) to give 34b (0.103 g, 86%). A solution of formaldehyde (0.081 g of a 38%, w/v, solution, 1.0 mmol) in MeOH (11 mL) was added to a stirred solution of 34b (0.097 g, 0.36 mmol), NaCNBH<sub>3</sub> (0.05 g, 0.80 mmol), and glacial acetic acid (0.103 mL, 1.8 mmol), in MeOH (11 mL) at 0 °C. The mixture was stirred at 0 °C for 2 h and then at room temperature for 1 h. Saturated K<sub>2</sub>CO<sub>3</sub> solution (5 mL) was added, the MeOH evaporated under reduced pressure, and the residue taken up into ethyl acetate (30 mL) and washed with brine  $(2\times)$ . The crude product was chromatographed on silica gel eluting with CH<sub>2</sub>Cl<sub>2</sub>/EtOH/NH<sub>3</sub> (40:8:1) to give **36b** (0.075 g, 70%). The hydrogen oxalate salt was prepared. 36b: mp 140-142 °C  $(MeOH/Et_2O)$ ; <sup>1</sup>H NMR  $(D_2O)$   $\delta$  1.54 (3H, t, J = 7.4 Hz, CH<sub>2</sub>CH<sub>3</sub>), 2.91 (6H, s, N(CH<sub>3</sub>)<sub>2</sub>), 3.21 and 3.47 (each 2H, each t, J = 7.4 Hz, 1-CH<sub>2</sub> and 2-CH<sub>2</sub>), 4.34 (2H, s, CH<sub>2</sub>), 4.64 (2H, q, J = 7.4 Hz,  $CH_2CH_3$ ), 7.17 (1H, dd, J = 1.5 and 8.4 Hz, 6'-CH), 7.33 (1H, s, 2'-CH), 7.48 (1H, d, J = 8.4 Hz, 7'-CH), 7.59 (1H, s, 4'-CH). Anal. (C16H22N6 (COOH)2) C, H, N.

The following tetrazoles were prepared using the general procedure described for 36b.

N,N-Dimethyl-2-[5-[(1-methyltetrazol-5-yl)methyl]-1Hindol-3-yl]ethylamine hydrogen oxalate (35a): mp 176-177 °C (MeOH/Et<sub>2</sub>O); <sup>1</sup>H NMR (D<sub>2</sub>O)  $\delta$  2.91 (6H, s, N(CH<sub>3</sub>)<sub>2</sub>), 3.21 and 3.40 (each 2H, each t, J = 7.4 Hz,  $1-CH_2$  and  $2-CH_2$ ), 4.00 (3H, s, CH<sub>3</sub>), 4.43 (2H, s, CH<sub>2</sub>), 7.13 (1H, dd, J = 1.5 and 8.4 Hz, 6'-CH), 7.35 (1H, s, 2'-CH), 7.50 (1H, d, J = 8.4 Hz, 7'-CH), 7.54 (1H, s, 4'-CH). Anal.  $(C_{15}H_{20}N_6 \cdot (COOH)_2)$  C, H, N.

N,N-Dimethyl-2-[5-[(2-methyltetrazol-5-yl)methyl]-1Hindol-3-yl]ethylamine hydrogen oxalate (35b): mp 185-186 °C (MeOH/Et<sub>2</sub>O); <sup>1</sup>H NMR (D<sub>2</sub>O)  $\delta$  2.91 (6H, s, N(CH<sub>3</sub>)<sub>2</sub>), 3.21 and 3.47 (each 2H, each t, J = 7.4 Hz, 1-CH<sub>2</sub> and 2-CH<sub>2</sub>), 4.30 (3H, s, CH<sub>3</sub>), 4.34 (2H, s, CH<sub>2</sub>), 7.17 (1H, dd, J = 1.5 and 8.4 Hz, 6'-CH), 7.33 (1H, s, 2'-CH), 7.48 (1H, d, J = 8.4 Hz, 7'-CH), 7.59 (1H, s, 4'-CH). Anal. (C<sub>15</sub>H<sub>20</sub>N<sub>6</sub>·(COOH)<sub>2</sub>) C, H, Ν

N.N-Dimethyl-2-[5-[(1-ethyltetrazol-5-yl)methyl]-1Hindol-3-yl]ethylamine hydrogen oxalate (36a): mp 175-178 °C (MeOH/Et<sub>2</sub>O); <sup>1</sup>H NMR (D<sub>2</sub>O)  $\delta$  1.32 (3H, t, J = 7.4 Hz, CH<sub>2</sub>CH<sub>3</sub>), 2.90 (6H, s, N(CH<sub>3</sub>)<sub>2</sub>), 3.21 and 3.46 (each 2H, each t, J = 7.4 Hz, 1-CH<sub>2</sub> and 2-CH<sub>2</sub>), 4.38 (2H, q, J = 7.4 Hz,  $CH_2CH_3$ , 4.47 (2H, s,  $CH_2$ ), 7.14 (1H, dd, J = 1.5 and 8.4 Hz, 6'-CH), 7.35 (1H, s, 2'-CH), 7.50 (1H, d, J = 8.4 Hz, 7'-CH), 7.53 (1H, s, 4'-CH). Anal. (C<sub>16</sub>H<sub>22</sub>N<sub>6</sub>·1.05(COOH)<sub>2</sub>) C, H, N.

Biochemical Methods. Biochemical methods for both the binding and functional experiments have previously been described.21

**Pharmacokinetic Methods.** For each compound studied, each one of a number of rats (36-72 rats having been deprived)of food overnight and typically weighing between 175 and 250 g) was allocated to one of a range of blood sampling time points, the latest point being between 4 and 8 h after drug administration. Six rats were allocated to each time point, of which three were dosed intravenously (iv) and three were dosed orally. The dose was always 3 mg of free base (or free base equivalent)/kg of body weight. Each rat was anesthetized with isoflurane at its preselected blood sampling time point, and a large sample of blood (>5 mL) was drawn into a lithium heparin blood tube by cardiac puncture. Plasma was separated from the blood by centrifugation and stored at -20 °C until analysis. Extracts of plasma were prepared either by liquid-liquid or solid phase extraction and analyzed by reversed phase HPLC employing UV or fluorescence detection. The average plasma concentration value at each dose route/ sampling time combination was calculated from the three individual data points, and pharmacokinetic parameters were calculated from the mean data by standard model-independent methods.

Acknowledgment. We thank Dr. Richard Herbert for NOE experiments, our colleagues in the analytical group for  $\log D$  determinations, and Mrs. E. Brawn for typing the manuscript.

Supplementary Material Available: Table of microanalytical data for novel compounds (2 pages). Ordering information is given on any current masthead page.

#### References

- (1) This work was presented in part at the Groupe D'etude Structure Activite XXIII, Annecy, France, May 1993, the 7th RSC-SCI Medicinal Chemistry Symposium, Cambridge, U.K., Sept. 1993 (P15), and the 208th National ACS Meeting, Washington, DC, Aug. 1994 (MEDI 180).
- Branchek, T. More Serotonin Receptors? Curr. Biol. 1993, 3 (5), (2)315 - 317
- 315-317.
  (a) Beer, M. S.; Middlemiss, D. N.; McAllister, G. 5-HT<sub>1-like</sub> Receptors: Six Down and Still Counting. Trends Pharmacol. Sci.
  1993, 14, 228-231. (b) Hartig, P. R.; Adham, N.; Zgombick, J.; Weinshank, R.; Branchek, T. Molecular Biology of the 5-HT<sub>1</sub> Receptor Subfamily. Drug Dev. Res. 1992, 26, 215-224. (c) Weinshank, R. L.; Zgombick, J. M.; Macchi, M.; Branchek, T. A.; Hartig, P. R. Human Serotonin 1D Receptor is Encoded by a Subfamily of Two Distinct Genes: 5-HT<sub>1Da</sub> and 5-HT<sub>1Ds</sub>. Proc. Natl. Acad. Sci. U.S.A. 1992, 89, 3630-3634. (d) Hartig, P. R.; Branchek, T. A.; Weinshank, R. L. A Subfamily of 5-HT<sub>1D</sub> Receptor Genes. Trends Pharmacol. Sci. 1992, 13, 152-159. (e) McAllister, G.; Charlesworth, A.; Snodin, C.; Beer, M. S.; Noble (3)McAllister, G.; Charlesworth, A.; Snodin, C., Beer, M. S.; Noble,

- A. J.; Middlemiss, D. N.; Iversen, L. L.; Whiting, P. Molecular Cloning of a Serotonin Receptor From Human Brain (5-HT<sub>1E</sub>) : A Fifth 5-HT1.like Subtype. Proc. Natl. Acad. Sci. U.S.A. 1992, 89, 5517-5521. (f) Adham, N.; Kao, H.-T.; Schechter, L. E.; Bard, J.; Olsen, M.; Urquhart, D.; Durkin, M.; Hartig, P. R.; Weinshank, R. L.; Branchek, T. A. Cloning of Another Human Serotonin Receptor  $(5-HT_{1F})$ : A Fifth 5-HT<sub>1</sub> Receptor Subtype Coupled to the Inhibition of Adenylate Cyclase. Proc. Natl. Acad. Sci. U.S.A. 1993, 90, 408-412.
- (4) Bockaert, J.; Fozard, J. R.; Dumuis, A.; Clarke, D. E. The 5-HT<sub>4</sub> Receptor. A Place in the Sun. Trends Pharmacol. Sci. 1992, 13, 141 - 145
- (5) Matthes, H.; Boschert, U.; Nourdine, A.; Grailhe, R.; Plassat, J.-L.; Muscatelli, F.; Mattei, M.-G.; Hen, R. Mouse 5-Hydroxytryptamine<sub>5A</sub> and 5-Hydroxytryptamine<sub>5B</sub> Receptors Define a New Family of Serotonin Receptors : Cloning, Func-tional Expression, and Chromosomal Localization. Mol. Phar-
- (6) Monsma, F. J., Jr.; Shen, Y.; Ward, R. P.; Hamblin, M. W.; Sibley, D. R. Cloning and Expression of a Novel Serotonin Receptor with High Affinity for Tricyclic Psychotropic Drugs. Mol. Pharmacol.
- (a) Shen, Y.; Monsma, F. J., Jr.; Metcalf, M. A.; Jose, P. A.; Hamblin, M. W.; Sibley, D. R. Molecular Cloning and Expression of a 5-Hydroxytryptamine<sub>7</sub> Serotonin Receptor Subtype. J. Biol. (7)Chem. 1993, 268 (24), 18200-18204. (b) Bard, J. A.; Zgombick, J.; Adham, N.; Vayasse, P.; Branchek, T. A.; Weinshank, R. L. Cloning of a Novel Human Serotonin Receptor (5-HT<sub>7</sub>) Positively Linked to Adenylate Cyclase. J. Biol. Chem. **1993**, 268 (31), 23422 - 23426
- (8) Plassat, J.-L.; Amlaiky, N.; Hen, R. Molecular Cloning of a Mammalian Serotonin Receptor That Activates Adenylate Cy-clase. Mol. Pharmacol. 1993, 44, 229-236.
- Humphrey, P. P. A.; Hartig, P.; Hoyer, D. A Proposed New Nomenclature for 5-HT Receptors. *Trends Pharmacol. Sci.* 1993, (9)14, 233–236.
- (10) Glennon, R. A.; Westkaemper, R. B. 5-HT<sub>1D</sub> Receptors: A Serotonin Receptor Population for the 1990s. Drug News Perpect. 1993, 6 (6), 390-405.
- (11) Heuring, R. E.; Peroutka, S. J. Characterisation of a Novel <sup>3</sup>H-5-Hydroxytryptamine Binding Site Subtype in Bovine Brain Membranes. J. Neurosci. 1987, 7 (3), 894–903. (12) Nowak, H. P.; Mahle, C. D.; Yocca, F. D. [<sup>3</sup>H]-5-carboxamido-
- tryptamine labels 5-HT<sub>1D</sub> binding sites in bovine substantia nigra. Br. J. Pharmacol. 1993, 109, 1206-1211.
  (13) (a) Feniuk, W.; Humphrey, P. P. A. The Development of a Highly
- Selective 5-HT<sub>1</sub> Receptor Agonist, Sumatriptan, for the Treatment of Migraine. Drug Dev. Res. 1992, 26, 235-240. (b) Dechant, K. L.; Clissold, S. P. Sumatriptan : A Review of its Pharmacodynamic and Pharmacokinetic Properties, and Therapeutic Efficacy in the Acute Treatment of Migraine and Cluster Headache. Drugs 1992, 43 (5), 776–798. (14) Glennon, R. A.; Ismaiel, A. M.; Chaurasia, C.; Titeler, M. 5-HT<sub>1D</sub>
- Serotonin Receptors : Results of a Structure-Affinity Investigation. Drug Dev. Res. 1991, 22, 25–36.
- (15) King, F. D.; Brown, M. A.; Gaster, M. L.; Kaumann, A. J.; Medhurst, A. D.; Parker, S. G.; Parsons, A. A.; Patch, T. L.; Raval, P. (±) 3-Amino-6-carboxamido-1,2,3-tetrahydrocarbazole: A Conformationally Restricted Analogue of 5-Carboxamidotryptamine with Selectivity for the Serotonin 5-HT<sub>1D</sub> Receptor. J. Med. Chem. 1993, 36, 1918-1919.
- (16) (a) Glen, R. C.; Hill, A. P.; Martin, G. R.; Robertson, A. D. Molecular Design of 5-HT<sub>1D</sub> Agonists for the Acute Treatment of Migraine. Headache 1994, 34, 307. (b) Hill, A. P.; Hyde, R. M.; Robertson, A. D.; Wollard, P. M.; Glen, R. C.; Martin, G. R. Oral Delivery of 5-HT1D Receptor Agonists: Towards the Discovery of 311C90, a Novel Anti-Migraine Agent. Headache 1994, 34. 308.
- (17) Macor, J. E.; Blank, D. H.; Post, R. J.; Ryan, K. The Synthesis of a Conformationally Restricted Analog of the Anti-Migraine Drug Sumatriptan. Tetrahedron Lett. 1992, 33 (52), 8011-8014.
- Ward, T. J. 5-HT<sub>1</sub>-Like and 5-HT<sub>1D</sub> Agonists as Treatments for Migraine. *Curr. Opin. Ther. Pat.* **1993**, 417–423.
   (a) Feniuk, W.; Humphrey, P. P. A.; Perren, J. J. The Selective
- Carotid Arterial Vasoconstrictor Action of GR 43175 in Anaesthetized Dogs. Br. J. Pharmacol. 1989, 96, 83-90. (b) Saxena, P. R.; Ferrari, M. D. 5-HT<sub>1-Like</sub> Receptor Agonists and the Pathophysiology of Migraine. Trends Pharmacol. Sci. 1989, 10, 200 - 204
- (20) (a) Gabriella Buzzi, M.; Moskowitz, M. A. The Antimigraine Drug, Sumatriptan (GR 43175), Selectively Blocks Neurogenic Plasma Extravasation from Blood Vessels in Dura Mater. Br. J. Pharmacol. 1990, 99, 202-206. (b) Moskowitz, M. A. Neurogenic versus Vascular Mechanisms of Sumatriptan and Ergot Alkaloids in Migraine. Trends Pharmacol. Sci. **1992**, 13, 307-311.
- (21) Street, L. J.; Baker, R.; Castro, J. L.; Chambers, M. S.; Guiblin, A. R.; Hobbs, S. C.; Matassa, V. G.; Reeve, A. J.; Beer, M. S.; Middlemiss, D. N.; Noble, A. J.; Stanton, J. A.; Scholey, K.;

Hargreaves, R. J. Synthesis and Serotonergic Activity of 5-(Oxadiazolyl)tryptamines : Potent Agonists for 5-HT<sub>1D</sub> Receptors. J. Med. Chem. 1993, 36, 1529-1538.

- (22) Castro, J. L.; Matassa, V. G.; Broughton, H. B.; Mosley, R. T.; Street, L. J.; Baker, R. Synthesis, Biological Activity and Electrostatic Properties of 3-[2-(Dimethylamino)ethyl]-5-[(3amino-1,2,4-thiadiazol-5-yl)methyl]-1H-indole, a Novel 5-HT1D Receptor Agonist. Bioorg. Med. Chem. Lett. 1993, 3 (6), 993-997.
- (23) Beer, M. S.; Stanton, J. A.; Bevan, Y.; Heald, A.; Reeve, A. J.; Street, L. J.; Matassa, V. G.; Hargreaves, R. J.; Middlemiss, D. N. L-694,247 : a Potent 5-HT<sub>1D</sub> Receptor Agonist. Br. J. Pharmacol. 1993, 110, 1196-1200.
- (24) A preliminary account of the profile of the triazole 10a was presented at the 3rd IUPHAR Satellite Meeting on Serotonin, Chicago, IL, July 1994.
- (a) Polya, J. B. 1,2,4-Triazoles. In Comprehensive Heterocyclic (25)Chemistry; Potts, K. T., Ed.; Pergamon Press: New York, 1984; Vol. 5, p 733. (b) This is the Grandberg modification of the Fischer Indole Synthesis. Robinson, B. The Fischer Indole Synthesis; John Wiley and Sons: New York, 1982; pp 487-495.
- (26) Fleming, I.; Pearce, A. Controlling the Outcome of a Carbocation-Initiated Cyclisation. J. Chem. Soc., Perkin Trans. 1 1981, 251-255.

- (27) Baker, R.; Matassa, V. G.; Street, L. J. Imidazole, Triazole and
- (27) Baker, N.; Matassa, V. G.; Street, L. J. Imidazole, Inazole and Tetrazole Derivatives. European Patent Appl. 0497512, 1992.
  (28) Chen, C.; Senanayake, C. H.; Bill, T. J.; Larsen, R. D.; Verho-even, T. R.; Reider, P. J. Improved Fischer-Indole Reaction for the Preparation of N,N-Dimethyltryptamines : Synthesis of Control of N,N-Dimethyltryptamines : Synthesis of L-695,894, a Potent 5-HT<sub>1D</sub> Receptor Agonist. J. Org. Chem. 1994, 59, 3738-3741.
- (29) Beer, M. S.; Stanton, J. A.; Bevan, Y.; Chauhan, N. S.; Middle-miss, D. N. An Investigation of the 5-HT<sub>1D</sub> Receptor Binding Affinity of 5-Hydroxytryptamine, 5-Carboxamidotryptamine and Sumatriptan in the Central Nervous System of Seven Species. Eur. J. Pharmacol. 1992, 213, 193-197.
- (30) Martin, G. R.; MacLennan, S. J. Analysis of the 5-HT Receptors in Rabbit Saphenous Vein Exemplifies the Problem of Using Exclusion Criteria for Receptor Classification. Naunyn-Schmiedeberg's Arch. Pharmacol. 1990, 342, 111-119.
- (31) Hargreaves, R. J.; Shepheard, S.; Beer, M. S.; Stanton, J. A.; Middlemiss, D. N.; et al. Unpublished results.
- (32) A detailed description of the pharmacokinetic profile of triazole 10a (MK-462) shall be the subject of a forthcoming publication. Hargreaves, R. J.; Graham, M. I.; Sohal, B.; et al. Unpublished results.

JM9500535